

## ORIGINAL ARTICLE

# Polymerase/nicking enzyme synergetic isothermal quadratic DNA machine and its application for one-step amplified biosensing of lead (II) ions at femtomole level and DNA methyltransferase

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Herein, we propose a novel and universal biosensing platform based on a polymerase-nicking enzyme synergetic isothermal quadratic DNA machine (ESQM). This platform tactfully integrates two signal amplification modules, strand displacement amplification (SDA) and nicking enzyme signal amplification (NESA), into a one-step system. A bifunctional DNA probe with a stem-loop structure was designed to be partly complementary to the SDA product and digested substrate of NESA for bridging SDA and NESA. ESQM can be performed by using only the enzymes and buffer involved in the SDA module. In the presence of a target, this DNA machine is activated to afford a high quadratic amplified signal. Using Pb<sup>2+</sup> and DNA adenine methylation (Dam) methyltransferase (MTase) as analytes, a sensitive biosensing platform is demonstrated. Low detection limits of 30 fM Pb<sup>2+</sup> and 0.05 U ml<sup>-1</sup> Dam MTase were achieved within a short assay time (40 min), which were each superior to those of most previously reported methods. This DNA machine exhibited high selectivity for Pb<sup>2+</sup>. Furthermore, the successful detection of complex environmental water samples demonstrated the applicability of the proposed strategy in real samples, holding great potential for its application in environmental monitoring, biomedical research and clinical diagnosis.

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## INTRODUCTION

Signal amplification technologies have a critical role in molecular biology research, environmental and food monitoring, forensic identification and clinical diagnosis. Since its creation in 1985, polymerase chain reaction has become the most well-known and widely used amplification technique.<sup>1</sup> However, this technique suffers from an intrinsic drawback, the requirement of precision thermal cycling between three different temperatures in a costly thermal cycler, limiting its popularization and application in point-of-care testing. On the other hand, various alternative isothermal amplification methods, such as rolling circle amplification (RCA),<sup>2,3</sup> strand displacement amplification (SDA),<sup>4</sup> loop-mediated isothermal amplification,<sup>5,6</sup> nicking enzyme signal amplification (NESA)<sup>7-9</sup> and the exponential amplification reaction (EXPAR)<sup>10</sup> have been proposed in the past two decades. These isothermal methods have been developed mainly based on some new findings in molecular biology concerning DNA synthesis and some accessory proteins together with their mimicking *in vitro* for nucleic acid amplification. Among these methods, EXPAR has been

widely used, which is attributed to its rapid amplification kinetics (several minutes) and exceedingly high amplification efficiency (10<sup>6</sup>- to 10<sup>9</sup>-fold). For example, Ye's group<sup>11</sup> and Zhang's group<sup>12,13</sup> have proposed several interesting methods for the highly sensitive detection of microRNA and protein based on EXPAR, respectively. Nevertheless, the EXPAR-based methods involve the weaknesses of early phase non-specific background amplification resulting from the binding of the polymerase to the single-stranded template.<sup>14</sup> Conversely, the linear amplification strategies such as RCA, SDA and NESA are much simpler than EXPAR and out of the non-specific background amplification. Though well established and widely applied, these linear amplification strategies still face challenges in the detection of trace targets because of their relative low sensitivities.

Recently, the cascade amplification strategies, combining two or more amplification techniques together, have attracted an explosion of interest due to their ability to integrate the merits of different methods. These approaches exhibit a comparable sensitivity to the EXPAR-based methods and can avoid the high background signal caused by

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non-specific amplification. In 2008, Xie and co-workers proposed a sensitive DNA biosensor based on cascade enzymatic signal amplification by combining RCA and NESAs.<sup>7</sup> Compared with the basic NESAs, this strategy significantly improved the detection sensitivity by nearly two orders of magnitude. Recently, several novel RCA-combined cascade amplification approaches have been successively developed for the high sensitive or ultrasensitive detection of nucleic acids, micro-RNA and cancer cells.<sup>15–17</sup> However, all of these methods suffer from lengthy assay times (several hours) with multi-step and complex reactions. In addition, SDA was also combined with other amplification techniques to design a cascade amplification system, which is attributed to its simplicity over RCA. For instance, Willner and colleagues proposed a promising DNA machine in a one-step reaction based on integrating SDA with an  $Mg^{2+}$ -dependent DNase-assisted recycling cleavage reaction.<sup>18</sup> Furthermore, these researchers developed an improved system that was extended to the amplified detection of cocaine.<sup>19</sup> Unfortunately, a long time (more than 8 h) was required. Moreover, the substrate for the  $Mg^{2+}$ -dependent DNase, as a signal reporter, contains a ribonucleotide that is easily degradable and requires strict operating conditions, which might cause false positive results, especially within a lengthy detection time. Recently, a two-stage isothermal amplification assay was proposed by introducing SDA into NESAs with a short linear quencher-fluorophore DNA probe.<sup>20</sup> This assay improved the detection sensitivity by as much as two orders of magnitude. Nevertheless, this method was cursorily constructed by the straightforward addition of SDA and NESAs, leading to a multi-enzyme (three enzymes) and multi-step (four steps) reaction system with a long assay time (more than 2 h). Therefore, the development of one-step, rapid and simple cascade amplification strategies for ultrasensitive detection remains an urgent need.

Inspired by the dramatic finding that the reaction system of SDA contains the enzyme and buffer for the NESAs reaction, herein, we construct a novel biosensor based on the polymerase-nicking enzyme synergetic isothermal quadratic DNA machine (ESQM) by integrating the SDA module with the NESAs module into a one-step and simple biosensing system. ESQM can be performed in a reaction system of the basic SDA without the additional enzyme and buffer involved in NESAs. In this DNA machine, a stem-loop structure bifunctional DNA probe (hairpin probe) is specially designed to be complementary with the SDA product and substrate of NESAs simultaneously, acting as a ‘bridge’ to connect the SDA module and NESAs module. Using a commercially synthesized oligonucleotide as a trigger, a high signal-to-noise ratio (18-fold) was obtained within 30 min using our quadratic amplification DNA machine. Then, the detection of  $Pb^{2+}$  was performed using ESQM combined with  $Pb^{2+}$ -dependent GR-5 DNase as the recognition probe. An ultralow detection limit down to 30 fM  $Pb^{2+}$  was achieved. Complex environmental water samples were successfully detected by the proposed biosensor. The results are in good agreement with those obtained using inductively coupled plasma mass spectrometry (ICP-MS), demonstrating the great potential of this biosensing system. Furthermore, this amplification DNA machine has also been readily expanded for the detection of DNA methyltransferase (MTase) activity and its inhibition by several anticancer or antimicrobial drugs.

## EXPERIMENTAL PROCEDURE

### Materials and reagents

Lead (II) acetate trihydrate and other metal salts,  $Hg(NO_3)_2$ ,  $MgCl_2$ ,  $MnCl_2$ ,  $CaCl_2$ ,  $SnCl_2$ ,  $FeCl_3$ ,  $CdCl_2$ ,  $CoCl_2$  and  $ZnCl_2$ , were purchased from Sigma-Aldrich (St Louis, MO, USA). Klenow fragment ( $exo^-$ ), Nt.BbvCI, DNA adenine methylation (Dam) MTase and the corresponding buffers were

obtained from New England Biolabs Ltd. (Beijing, China). All the oligonucleotides were synthesized by Sangon Biotechnology Co., Ltd. (Shanghai, China). The sequences are listed in Supplementary Table S1. All the other solutions were prepared and diluted with Milli-Q water (resistance > 18.2 M $\Omega$ ).

### $Pb^{2+}$ detection based on ESQM

To detect  $Pb^{2+}$ , various concentrations of  $Pb^{2+}$  were added to 20  $\mu$ l of 1 $\times$  NEBuffer 2 containing 10 nM DNase, 10 nM SDA template, 50 nM hairpin probe, 250 nM molecular beacon (MB), 2.5 units of Klenow Fragment, 0.8 units of Nt.BbvCI and 0.2 mM dNTPs. Subsequently, the solutions were incubated at 37  $^\circ$ C for 40 min before the fluorescence measurement. The river water samples were first filtered through a 0.2- $\mu$ m membrane to remove the insoluble substance and were then treated by 5%  $HNO_3$  and 10 mM  $Mg^{2+}$  before testing under the same conditions described above. A Thermo X Series 2 ICP-MS (Thermo Fisher Scientific, Shanghai, China) was used in this work.

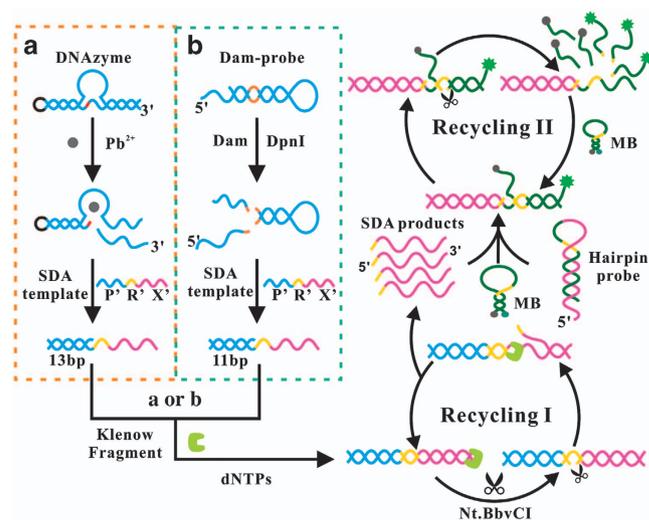
### Application for Dam MTase activity assay

The experiments were performed in 20  $\mu$ l 1 $\times$ NEBuffer 2 containing a 25-nM Dam-probe, 10 nM SDA template, 50 nM hairpin probe, 250 nM MB, 160  $\mu$ M SAM, 2 units of *DpnI*, 0.2 mM dNTPs, 0.8 units of Nt.BbvCI and 2.5 units of Klenow Fragment. After the addition of various concentrations of Dam MTase, the solutions were incubated at 37  $^\circ$ C for 40 min. Then, fluorescence emission spectra were recorded as described above.

## RESULTS AND DISCUSSION

### Principle of ESQM-based biosensor

The principle of the ESQM-based biosensor is outlined in Figure 1. Three main components, SDA template, bifunctional hairpin probe and MB (digested substrate of NESAs) are employed. The SDA template consists of a primer-binding domain ( $P'$ , blue), an amplification domain ( $X'$ , red violet) and a recognition sequence ( $R'$ , yellow) for nicking enzyme Nt.BbvCI that recognizes double-strand DNA and cleaves only one strand. The bifunctional hairpin probe containing a stem-loop structure is specially designed to be partly complementary to the SDA product and digested substrate of NESAs, respectively. This probe is used as a ‘bridge’ to connect the SDA module and NESAs



**Figure 1** Schematic illustration of ESQM and its application for  $Pb^{2+}$  and Dam MTase detection. (a)  $Pb^{2+}$  activates GR-5 DNase for the cleavage reaction at rA (highlighted in red), which generates a primer to trigger ESQM. (b) ESQM is initiated by the primer produced via the Dam MTase and *DpnI*-coupled reaction. Recycling I shows the SDA process of ESQM and Recycling II depicts the NESAs process induced by SDA products and the hairpin probe, leading to a quadratic amplified signal.

module. As the digested substrate of NESAs, the MB-containing recognition sequence for Nt.BbvCI also serves as a signal unit and is designed to be a stem-loop structure labeled with a fluorophore and quencher at its 5' and 3' end, respectively. The fluorescence signal is quenched due to the fluorescence resonance energy transfer reaction between the fluorophore and the quencher.

When the target is present (A or B), ESQM is activated. In detail, the extension reaction in the SDA module is initiated with the help of the DNA polymerase Klenow Fragment and dNTPs, producing the double-stranded SDA template containing the full recognition site for Nt.BbvCI. Following the nicking reaction by Nt.BbvCI, a new extension will continue with the displacement and release of the single-strand (ss) DNA SDA product. In this way (Recycling I), thousands of SDA products are generated, which open the hairpin probe with complementary hybridization as the trigger for NESAs. Then, the opened-hairpin probe can hybridize to MB, forming a DNA duplex with the full recognition site for Nt.BbvCI. After the nicking reaction of MB, the separation of the fluorophore from the quencher occurs as well as the release of the opened-hairpin probe. The disassociated opened-hairpin probe can hybridize with another un-cleaved MB and trigger the second cycle of nicking, leading to an amplified detection signal (Recycling II). Eventually, each target can undergo a quadratic amplification signal by ESQM. Note that the sensing process is performed in a one-step reaction at 37 °C. Furthermore, the nicking enzyme and reaction buffer involved in SDA are also suitable for NESAs. All of these merits are superior to those of some reported methods.<sup>7,17,20</sup>

In theory, we can calculate the recycling process of the linear and quadratic amplification system as detailed in Supplementary Table S2. Their signal equations are described as  $Y_1 = N$  for the linear amplification mode (basic SDA, see Supplementary Figure S1) and  $Y_2 = 0.5 N(N - 1)$  for the quadratic amplification mode (ESQM), where  $Y_1$  and  $Y_2$  represent the signal of the linear and quadratic amplification mode, respectively, and  $N$  is the positive integer.  $Y_2$  is always greater than  $Y_1$  when  $N > 3$  (meaning a sufficient reaction time). To confirm the amplification efficiency of our proposed DNA machine, a series of experiments was performed for comparison with the basic SDA without the integration of the NESAs module. As demonstrated in the inset of Figure 2, the background fluorescence intensity of ESQM in the absence of SDA primer (curve c) slightly increased compared with that of the basic SDA (curve d). This result should be due to the damage of the stem-loop structure of MB by partial hybridization with the hairpin probe. Fortunately, when the SDA primer (10 nM) is present, the fluorescence signal of ESQM (curve a) is much higher than that of the basic SDA process (curve b). A high signal enhancement of 18.0-fold was observed in the quadratic amplified strategy, whereas only a 10.1-fold signal increase was achieved by the basic SDA. These results, in accordance with our above theoretical analysis, indicate that the proposed DNA machine can provide a higher amplified signal by integrating the SDA module with the NESAs module.

#### Optimization of sensing conditions

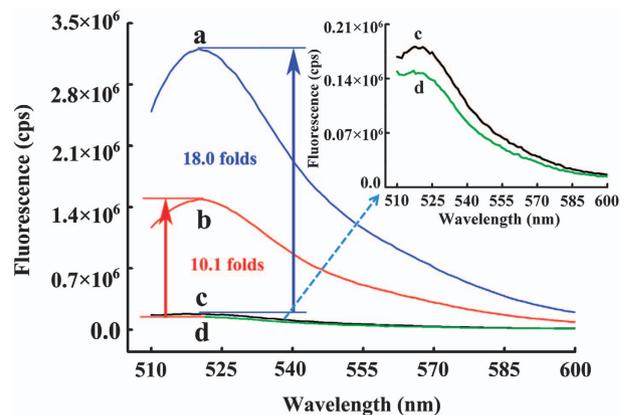
As the 'bridge' between the SDA module and NESAs module, the hairpin probe is specifically designed in our strategy. Its structure is very important for the sensing performance. In detail, this probe should be easily opened by the SDA product and hybridize with MB to trigger the NESAs process rapidly in the presence of a target. Conversely, when the target is absent, the hairpin probe should be stabilized and fail to hybridize with MB because of the strong intramolecular hybridization with the long stem, yielding a low

background signal. To obtain the best sensing performance, the stem length of the hairpin probe was investigated. As presented in Supplementary Table S1, there are three different hairpin probes (H1, H2 and H3), each including 8 bp, 10 bp and 12-bp in the stem region. Their potential secondary structures and the corresponding Gibbs-free energy are shown in Supplementary Figure S2. Their fluorescence responses are shown in Supplementary Figure S3. In the presence of a target, decreased fluorescence intensity was observed with the lengthening of the stem, which revealed that the hairpin probe with short stem can be more easily opened by the SDA product to produce a higher signal. Unfortunately, higher background fluorescence also occurred with the shorter stem hairpin probe, mainly due to the competing hybridization between MB and the hairpin probe. While H1 provided the highest fluorescence intensity, H2 was used in the experiments because of its highest signal-to-noise ratio. Subsequently, the molar ratio of the hairpin probe to MB (concentration is fixed at 250 nM) and the enzyme concentrations were optimized. The results are presented in Figures 3a–c. On the basis of the highest  $F/F_0$  value, a 1:5 molar ratio of the hairpin probe to MB, 0.8 units of Nt.BbvCI and 2.5 units of Klenow Fragment were employed for the following experiments.

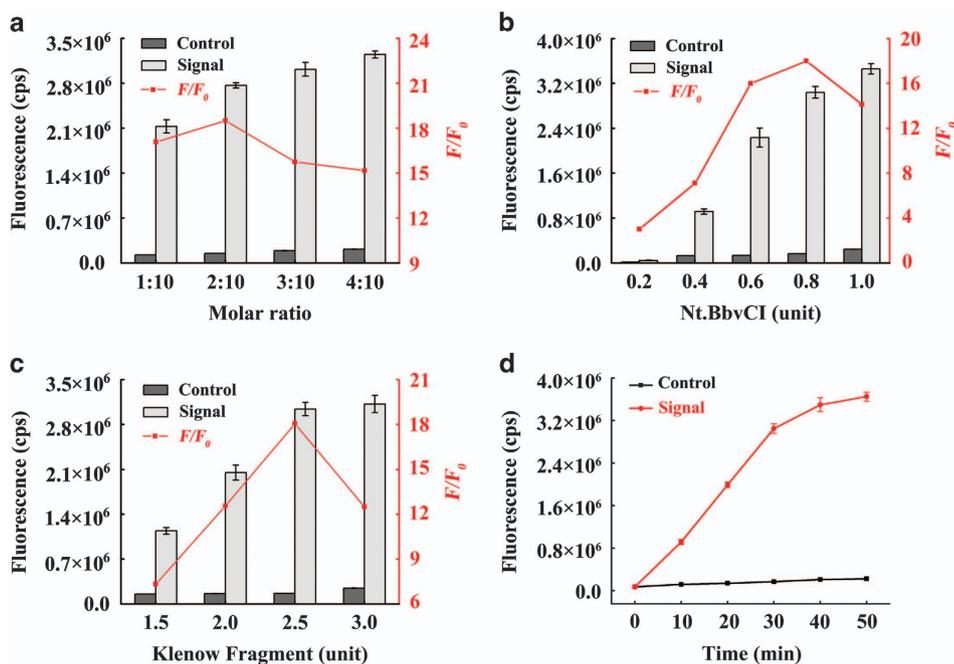
In addition, the effect of the reaction time was also investigated. As depicted in Figure 3d, with the prolongation of the reaction time, the fluorescence intensity in the presence of the SDA primer rapidly increased, indicating that the recycling cleavage of MB by the quadratic amplification integrating SDA with NESAs was triggered. The intensity nearly approached steady state after incubation for 50 min. Note that a slow fluorescence enhancement was also observed in the absence of the SDA primer. This background fluorescence signal mainly results from the competing hybridization between the hairpin probe and MB, which damaged the stem-loop structure of MB to produce a fluorescence signal. To obtain the best sensing performance, 30 min was selected because of the highest fluorescence enhancement. This assay time and the high signal-to-noise ratio (18-fold) are superior to those of the previously reported methods based on cascade amplification.<sup>21–25</sup>

#### Analytical performance for $Pb^{2+}$ detection based on ESQM

For the detection of  $Pb^{2+}$ , a DNAzyme (GR-5 DNAzyme) is involved in the proposed biosensor as a  $Pb^{2+}$  recognition unit. GR-5 DNAzyme is a functionalized nucleic acid with catalytic activity, cleaving the



**Figure 2** The fluorescence responses of the proposed sensing system (a) and basic SDA system (b) in the presence of 10 nM SDA primer, with corresponding backgrounds without SDA primer (c, d), respectively. Inset: Fluorescence spectra of (c) and (d).



**Figure 3** Optimization of the experimental conditions. The effect of the molar ratio of the hairpin probe to MB (a), Nt.BbvCI concentration (b) and Klenow Fragment concentration (c) on the fluorescence response of the DNA machine.  $F_0$  and  $F$  represent the fluorescence intensity in the absence and presence of 10 nM SDA primer, respectively. (d) The time-dependent fluorescence response with 10 nM SDA primer over background fluorescence. The error bar indicates the standard deviation of three independent experiments.

oligonucleotide substrate at the ribonucleotide (rA) recognition position in the presence of  $Pb^{2+}$ . In the absence of  $Pb^{2+}$ , the GR-5 DNAzyme exhibits no activity, and the substrate strand of DNAzyme is not cleaved. There is no primer existing to trigger the DNA machine. Because Nt.BbvCI is disabled to cleave the ssDNA, no fluorescence enhancement is observed. However, when  $Pb^{2+}$  is present, the recognition unit GR-5 DNAzyme performs the catalytic reaction for the hydrolytic cleavage of the substrate strand at rA (Figure 1a). Due to the low  $T_m$  ( $\sim 26^\circ\text{C}$ ), an ssDNA (13 mer) is released at the reaction temperature ( $37^\circ\text{C}$ ). Then, the released ssDNA hybridizes with the SDA template and acts as a primer to initiate the SDA process in the assistance of DNA polymerase and dNTPs. Finally, each  $Pb^{2+}$  can induce a quadratic amplified signal by ESQM as discussed above.

To obtain the best sensing performance, several important assay conditions including the concentration of DNAzyme and the reaction time were studied, and the results are presented in Figure 4. On the basis of the highest value of  $F/F_0$ , 10 nM DNAzyme and a 40-min reaction time were selected. Under the optimized conditions, gel-electrophoresis experiments were employed to demonstrate  $Pb^{2+}$  detection by the developed DNA machine. As shown in Figure 5a, no new product of cleaved MB was observed (lane a, in the absence of  $Pb^{2+}$ ). However, new bands of the cleaved MB were obviously present in the presence of  $Pb^{2+}$  (lane b), suggesting that  $Pb^{2+}$ -triggered quadratic amplification occurred. Then, a series of samples containing various  $Pb^{2+}$  concentrations were examined to evaluate the sensing performance of our sensing system. Figure 5b presents the fluorescence spectra in the presence of  $Pb^{2+}$  concentrations ranging from 0 to 50 nM. As observed, the fluorescence intensity gradually increased when the  $Pb^{2+}$  concentration was increased. With a correlation coefficient of  $R^2 = 0.9985$ , a linear relationship between the fluorescence intensity (518 nm) and logarithm of the  $Pb^{2+}$  concentration is

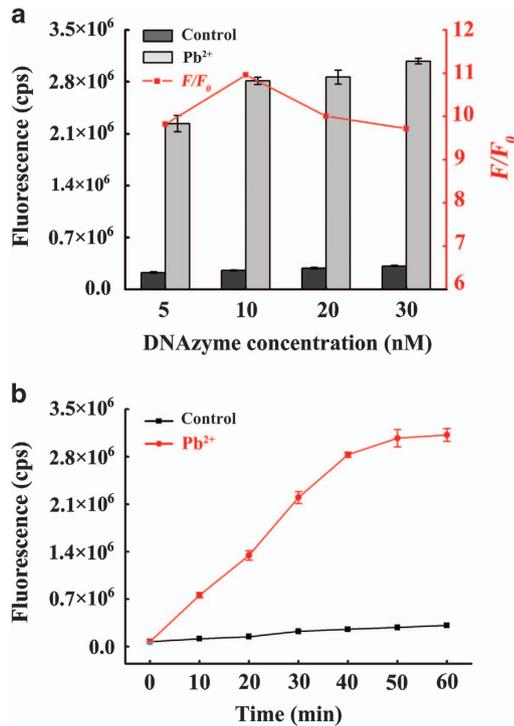
achieved (inset, Figure 5c). With a large dynamic range (6 orders of magnitude), a detection limit down to 30 fM was obtained based on three times standard deviation of the blank, which is at least 500-fold lower than those of previously reported fluorescence methods.<sup>22,26–29</sup> This sensitivity is also much higher than those of several electrochemical and electrochemiluminescence approaches<sup>24,25,30,31</sup> and is even comparable to the surface plasmon resonance-based biosensor.<sup>32</sup> The details are summarized in Table 1.

Control experiments with  $Pb^{2+}$  at various concentrations were also performed in the system only including basic SDA without the NES module (see Supplementary Figure S1). As demonstrated in Supplementary Figure S4, a detection limit of only 25 pM was obtained. This sensitivity is approximately three orders of magnitude lower than that of the proposed DNA machine integrating the SDA module with the NES module. Therefore, all these results revealed that a quadratic amplified signal and ultrahigh sensitivity for  $Pb^{2+}$  detection can be achieved with the amplified detection platform based on ESQM.

Selectivity was another important issue for the  $Pb^{2+}$  detection. Other environmentally relevant metal ions ( $Zn^{2+}$ ,  $Co^{2+}$ ,  $Ca^{2+}$ ,  $Hg^{2+}$ ,  $Cd^{2+}$ ,  $Fe^{3+}$ ,  $Mn^{2+}$ ,  $Sn^{2+}$  and  $Mg^{2+}$ ) were selected to investigate the selectivity of this assay. The fluorescent response of  $Pb^{2+}$  (10 nM) and other metal ions (100 nM) is shown in Figure 5d. Only  $Pb^{2+}$  triggered a significant fluorescence enhancement, whereas a very little increase in the fluorescent intensity was observed in the presence of the other interference ions. This finding indicated that the ESQM-based biosensing system exhibits a good specificity to  $Pb^{2+}$  over other competing metal ions. The excellent selectivity is due to the GR-5 DNAzyme, which is  $\sim 40\,000$  times more selective for  $Pb^{2+}$  against the most active interfering metal ion  $Zn^{2+}$ .<sup>33</sup> The 8–17 DNAzyme, another DNAzyme used for most  $Pb^{2+}$  biosensors, exhibits at most 160-fold selectivity against  $Zn^{2+}$ .<sup>33</sup>

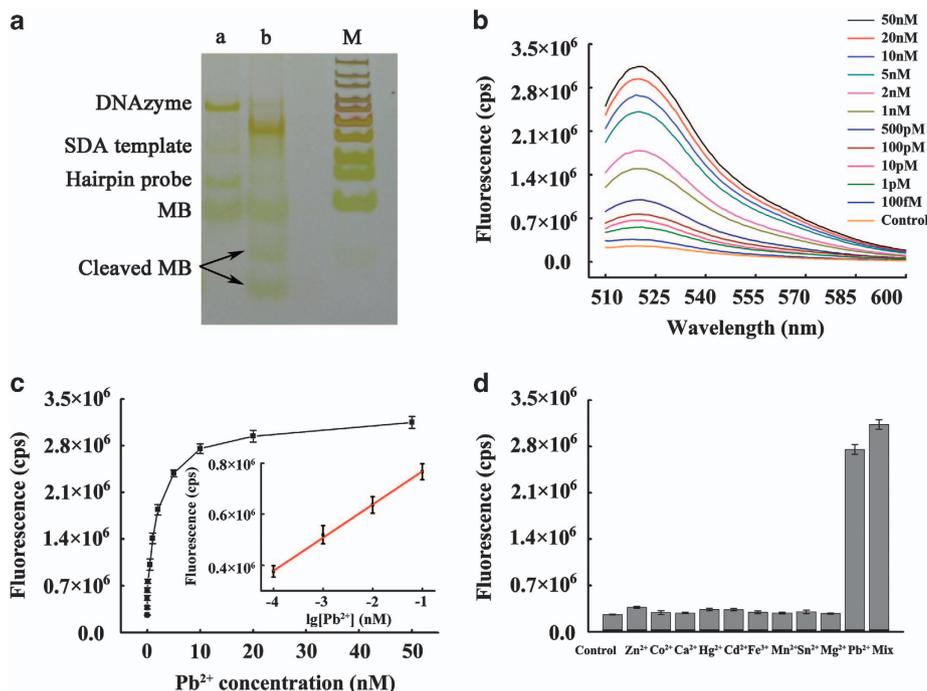
### Pb<sup>2+</sup> detection of water sample from Xi'an Chan River

The maximum contamination level of Pb<sup>2+</sup> in drinking water defined by the US Environmental Protection Agency is 72 nM, which is ~10<sup>6</sup>



**Figure 4** Optimization of DNAzyme concentration (a) and reaction time (b) for Pb<sup>2+</sup> detection.

fold higher than the detection limit of our sensing system. This requirement indicates that the proposed sensing system holds great potential for real water sample analysis of Pb<sup>2+</sup>. Acidification pretreatment of complex environmental samples is always used to reduce interference by organic matter and to convert metals associated with particulates to a form (usually a free metal) that can be detected by atomic absorption spectroscopy or ICP. Inspired by this procedure, the filtered river water was treated with an acidification process before analysis. Nitric acid was used due to the adequate digestion of most samples. As illustrated in Figure 6a, no obvious signal responses were observed for the ultrapure water samples (negative controls) treated with nitric acid by our biosensor. However, the filtered river water samples undergoing the acidification process exhibited significant Pb<sup>2+</sup>-dependent signal responses. In addition, the 5% (w/v) nitric acid resulted in the highest fluorescence intensity. Note that these water samples were diluted for the detection using our biosensor. The acidification time was also investigated, as depicted in Supplementary Figure S5A, and 1 h was selected as the signal reached a plateau effect. The above result of nitric acid-triggered Pb<sup>2+</sup> ‘output’ in the filtered river water may be attributed to the conversion of lead associated with particulate to free Pb<sup>2+</sup> and the release of free Pb<sup>2+</sup> from Pb<sup>2+</sup> coordination compounds under the acidic condition. Note that the GR-5 DNAzyme is selective for only free Pb<sup>2+</sup> rather than other forms, such as Pb<sup>2+</sup> coordination compounds. Furthermore, considering the dynamic equilibrium of the complexation reaction, we hypothesize that adding an overbalance of other metal ions as competitors may be helpful in displacing Pb<sup>2+</sup> from its coordination compounds. Mg<sup>2+</sup>, acting as a cofactor for most enzymes, is employed as a model competitor. As illustrated in Figure 6b, the introduction of a high Mg<sup>2+</sup> concentration (mM level) to the filtered river water leads to an

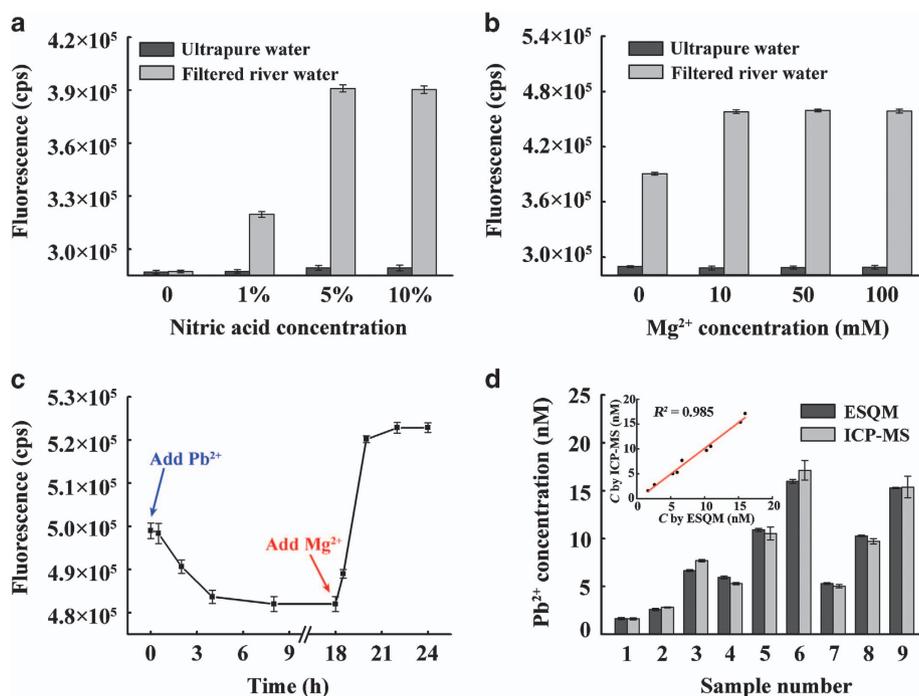


**Figure 5** (a) Non-denaturing PAGE analysis. Lane M, marker; lane a, Pb<sup>2+</sup> detection by ESQM in the absence of Pb<sup>2+</sup>; lane b, Pb<sup>2+</sup> detection by ESQM with 20 nM Pb<sup>2+</sup>. (b) Fluorescence spectra response to different concentrations of Pb<sup>2+</sup>. (c) Changes in fluorescence intensity at 518 nm upon the addition of various concentrations of Pb<sup>2+</sup>. The inset displays the linear correlation between the fluorescence intensity and the logarithm of the Pb<sup>2+</sup> concentration in the range of 100 fM to 100 pM. (d) Selectivity of the sensing strategy for Pb<sup>2+</sup> detection over other common interference metal ions and the mixture of these metal ions. The concentration of Pb<sup>2+</sup> is 10 nM, whereas the concentrations of other metal ions are each 100 nM.

**Table 1** Comparison of Pb<sup>2+</sup> detection methods

Method	Detection limit (pM)	Assay time (min)	Steps	Reference
Fluorescence detection based on ESAM	0.03	40	One step	Current method
Fluorescence detection based on lead(II)-driven DNA molecular device	20 000	120	One step	26
Amplified fluorescence detection by graphene and DNAzyme	300	20	One step	27
Amplified fluorescence detection based on catalytic MB	600	10	One step	28
Amplified fluorescence detection by exonuclease-aided DNA recycling	5	150	Two steps	22
Amplified fluorescence detection by isothermal strand displacement signal amplification	200	195	Multi-steps	29
Enzyme-free strip biosensor based on catalytic DNA circuit	10	105	Three steps	23
Amplified electrochemical assay by hybridization chain reaction	37	200	Three steps	24
ECL detection using DNAzyme	0.11	Not mentioned	One step	30
Electrochemical biosensor based on DNA functionalized gold nanoparticles	28	780	Three steps	31
Amplified electrochemical detection based on RCA	7.8	365	Multi-steps	25
Amplified SPR detection using gold nanoparticles	0.005	100	One step	32

Abbreviations: ECL, electrochemiluminescence; RCA, rolling circle amplification; SPR, surface plasmon resonance.



**Figure 6** Performance of the proposed biosensors in the analysis of environmental water samples. (a) The concentration-dependent effect of nitric acid on the Pb<sup>2+</sup>-triggered signal response. (b) The concentration-dependent effect of Mg<sup>2+</sup> on the Pb<sup>2+</sup>-triggered signal response. (c) Time-dependent Pb<sup>2+</sup> complexation and displacement in river water. (d) Determination of Pb<sup>2+</sup> in environmental water samples using the proposed ESQM biosensor and ICP-MS. Samples 1–9 represent drinking water samples spiked with 0, 1 and 5 nM Pb<sup>2+</sup>, Chan River water samples spiked with 0, 5 and 10 nM Pb<sup>2+</sup> and Ba River samples spiked with 0, 5 and 10 nM Pb<sup>2+</sup>, respectively. The inset shows the strong correlation between these two methods with a correlation coefficient of 0.985. *C* represents the Pb<sup>2+</sup> concentration.

increased Pb<sup>2+</sup>-dependent signal response, demonstrating our above hypothesis. In addition, the time of the Mg<sup>2+</sup> process was also optimized (Supplementary Figure S5B). To verify the complexation of Pb<sup>2+</sup> in the river water sample, filtered river water without acid or the Mg<sup>2+</sup> process was spiked with a standard Pb<sup>2+</sup> concentration. Time-dependent tests of this sample were performed after the addition of Pb<sup>2+</sup>. Figure 6c reveals that the signal of Pb<sup>2+</sup> decreases with time, implying the complexation of the added Pb<sup>2+</sup> by the complexing agent existing in the river water. Nevertheless, after adding excess Mg<sup>2+</sup> to the same sample, the Pb<sup>2+</sup>-triggered signal increases and is ultimately higher than that of the spiked Pb<sup>2+</sup> concentration, indicating the

successful displacement of both spiked Pb<sup>2+</sup> and intrinsic Pb<sup>2+</sup> from the coordination compounds.

Under the optimized condition of the nitric acid and Mg<sup>2+</sup> process, a linear relationship between the logarithm of the Pb<sup>2+</sup> concentration (0.1–100 pM) and fluorescence intensity is achieved, with the corresponding linear equation of  $I_F = 1.47 \lg[\text{Pb}^{2+}] + 4.67$ , where  $I_F$  represents the fluorescence intensity ( $\times 10^5$  cps). Then, filtered water samples from the Chan River treated with nitric acid and Mg<sup>2+</sup> were spiked with 0, 5 and 10 nM Pb<sup>2+</sup> and tested using the proposed ESQM biosensor. As depicted in Figure 6d, the Pb<sup>2+</sup> concentration in the Chan River water is 5.95 nM, as detected by ESQM. Satisfactory recoveries of 98.5 and 102.12% were obtained for the detection of

these spiked samples. These samples were also tested by ICP-MS to validate the  $Pb^{2+}$  concentrations, the result of which is presented in Figure 6d and the inset. The determined  $Pb^{2+}$  concentrations of the Chan River filtered water samples are consistent with the results obtained using our ESQM biosensor (correlation coefficient of 0.985). To further confirm the practical application of our biosensor, water samples from another river (the Ba River) (7–9 samples, Figure 6d) and drinking water samples (1–3 samples, Figure 6d) were also tested using our ESQM biosensor and ICP-MS with satisfactory results. These results clearly demonstrate that our proposed biosensor is capable of detecting  $Pb^{2+}$  in environmental water samples with excellent performance. In addition, the high sensitivity results in a high dilution factor for complex samples, which is significant for the decrease or even elimination of possible interference from different background compositions in these samples.

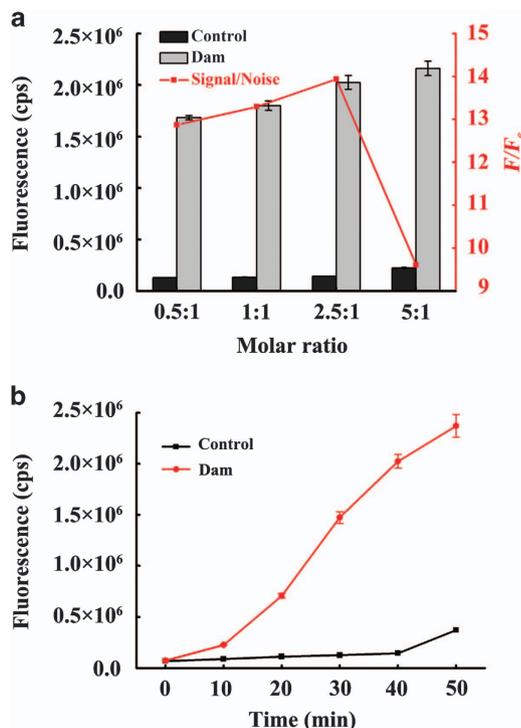
### Application of ESQM for the detection of DNA MTase

To demonstrate the versatility of the proposed strategy, we applied the developed DNA machine to construct another detection system for the assay of DNA MTase which is closely related to human disease such as cancer.<sup>34,35</sup> Herein, Dam MTase, a representative DNA MTase, was used as the model target. As illustrated in Figure 1b, a stem-loop structure Dam probe with a long stem was employed as the recognition unit for Dam MTase detection. In the presence of Dam MTase, the Dam probe is methylated, forming the substrate for *DpnI*. Subsequently, the cleavage of methylated Dam probe results in the release of a short ssDNA (11 mer) at the reaction temperature. This ssDNA is complementary to the SDA template, which can serve as an SDA primer to activate the DNA machine and generate an amplified fluorescence signal, as discussed above. Conversely, when Dam MTase is absent, *DpnI* cannot cut the Dam probe. The double helical structure of the Dam probe is maintained, and no primer is produced

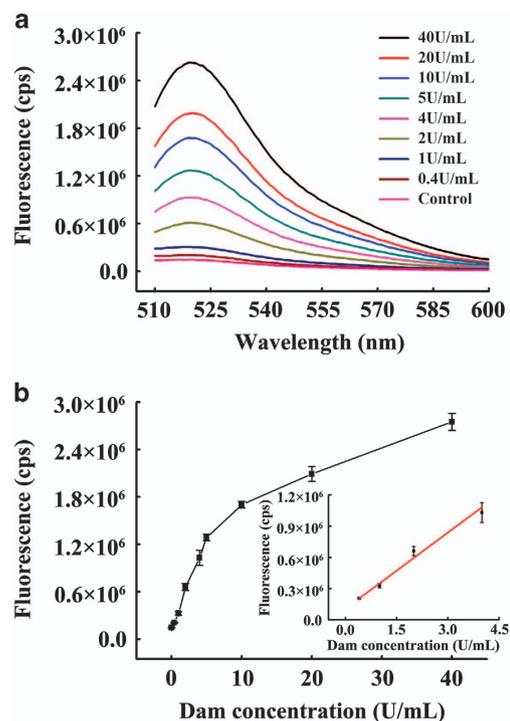
to trigger the DNA machine. Eventually, no obvious fluorescence signal is observed. Note that ESQM can be readily expanded for the detection of many other analytes, such as miRNA, ATP, cocaine and thrombin, with minor modification. For instance, only a new template is required for target miRNA to trigger ESQM. To design the sensing system for ATP, cocaine or thrombin, their aptamers are needed for target recognition and the trigger of ESQM. In all of these cases, the hairpin probe and MB do not need to be changed.

To obtain the best sensing performance, several conditions including the Dam probe concentration and reaction time were studied, the results of which are presented in Figure 7. It can be observed that both the 25-nM Dam probe and 40 min resulted in the highest signal-to-noise ratio, and these conditions were selected for the following experiments. Under these optimized assay conditions, various concentrations of Dam MTase were introduced, and the fluorescence spectra are shown in Figure 8a. The fluorescence intensity increased continuously with increasing Dam MTase concentration. Figure 8b shows the linear relationship between the fluorescence intensity and the Dam MTase concentration between 0.4 and 4 U ml<sup>-1</sup>. A low detection limit of 0.05 U ml<sup>-1</sup> was achieved, which is superior to that of most previously reported methods.<sup>36–38</sup> In addition, the high selectivity for Dam MTase against other enzymes, including M.SssI MTase and DNA ligase, was successfully demonstrated (Supplementary Figure S6).

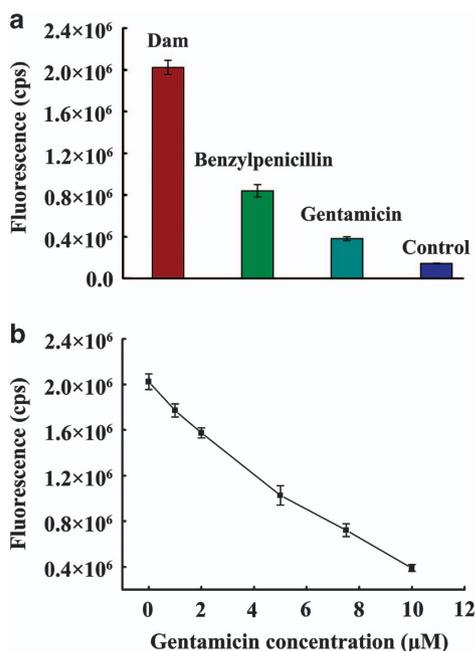
Numerous studies have demonstrated that Dam MTase has a vital role in several human diseases and bacterial virulence; thus, the evaluation of Dam MTase inhibition might contribute to important applications in antimicrobial drug development. In this work, several antimicrobial drugs were employed to investigate the inhibition of Dam MTase. The sensing system contains another three enzymes



**Figure 7** Effect of Dam probe concentration (a) and reaction time (b) on the performance of the Dam MTase assay.



**Figure 8** (a) Fluorescence spectra response to various concentrations of Dam MTase. (b) Changes in fluorescence intensity at 518 nm upon the addition of various Dam MTase concentrations. The inset displays the linear correlation between the fluorescence intensity and Dam MTase concentration.



**Figure 9** (a) Effect of benzylpenicillin (10 μM) and gentamicin (10 μM) on the activity of Dam MTase. (b) Inhibition of different concentrations of gentamicin on Dam MTase activity. The gentamicin concentrations were 0, 1, 2, 5, 7.5 and 10 μM.

(*DpnI*, Nt.BbvCI and Klenow Fragment); thus, control experiments (details are provided in Supplementary Information) were first performed to determine whether these drugs affect their activities. As depicted in Supplementary Figure S7, no obvious changes in the fluorescence intensity were observed in the absence and presence of these inhibitors, indicating that these drugs have no effect on their activities for concentrations of 10 μM. Thus, these concentrations of different inhibitors were used to investigate the inhibition of the Dam MTase activity. The fluorescence intensity in response to the inhibitors is shown in Figure 9a, and both inhibitors exhibited a significant inhibitory effect on Dam MTase. In addition, we further assessed the concentration-dependent inhibition of the Dam MTase activity using various gentamicin concentrations. Figure 9b demonstrates that the inhibition was gradually enhanced with increasing gentamicin concentration. Therefore, the proposed fluorescent strategy holds the potential to be a promising platform for the screening of antimicrobial drugs.

## CONCLUSIONS

In summary, we have developed a novel and universal biosensing platform based on the ESQM, integrating an SDA module and an NESA module into a one-step and simple sensing system. A bifunctional hairpin probe was specially designed as a bridge between these two isothermal signal amplification modules. This DNA machine was effectively implemented under the basic SDA reaction conditions without the introduction of additional enzymes or a complex reaction buffer. Using Pb<sup>2+</sup> as a model analyte, ultrasensitive detection with a detection limit of 30 fM was demonstrated in a short assay time (40 min), which is three orders of magnitude lower than that of a basic SDA assay. This sensitivity is also much higher than those of most reported fluorescence and electrochemical biosensors. Moreover, an excellent selectivity for Pb<sup>2+</sup> was achieved even in the presence of various interferential metal ions with high concentrations. The

satisfactory performance on river and drinking water samples demonstrated the potential applications of the proposed biosensor in complex environmental water samples. The versatility of the proposed sensing platform was also demonstrated by the successful detection of DNA MTase activity and its inhibition. As a universal sensing system, this DNA machine can be readily constructed to be an electrochemical or electrochemiluminescence biosensor by only changing the signal unit (MB in this work) to corresponding signal reporters (electrochemical MB or electrochemiluminescence MB).<sup>39,40</sup> Our ongoing work will demonstrate the applicability of this ESQM-based electrochemical biosensor combined with DNA nanostructure in the detection of cancer biomarkers such as microRNA and DNA methylation.

## CONFLICT OF INTEREST

The authors declare no conflict of interest.

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